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Molecular characteristics and virulence gene profiles of *Staphylococcus aureus* causing bloodstream infection



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ABSTRACT

Background: The rate of methicillin-resistant *Staphylococcus aureus* (MRSA) among the total of *S. aureus* isolates decreased to 35.3% in 2017 in China. It is unclear whether the molecular characteristics of *S. aureus* isolates have changed as the rate decreased.

Objective: This study aimed to investigate the molecular characteristics and virulence genes profile of *S. aureus* isolates causing bloodstream infection and analyze the correlation between the prevalence rates of the common sequence types and MRSA.

Methods: A total of 112 *S. aureus* strains from eight hospitals of four cities, including 32 MRSA isolates, were identified and evaluated through multilocus sequence typing, *spa* typing, and determination of virulence genes.

Results: Twenty-five STs were identified, of which ST5 (21.4%) was the most prevalent, whereas the prevalence of ST239 correlated with the rate of MRSA among all *S. aureus* isolates. Forty-six *spa* types were identified, of which t2460 (14.3%) was the most common. *clfa*, *hla*, *seb*, *fnbA* and *hly* were the prevailing virulence genes. 81.3% MRSA and 45.0% methicillin-sensitive *S. aureus* (MSSA) isolates harbored six or more tested virulence genes. ST5-t2460, seldom noted in bloodborne *S. aureus* isolates in China, was the most common clone. The prevalence of harboring six or more virulence genes in ST5-t2460 and ST188-t189 were 93.8% and 8.3%, respectively.

Conclusion: ST5-t2460 was the most common clone in *S. aureus* causing bloodstream infection followed by ST188-t189, which had never been noted in China before. Moreover, ST5-t2460 harbored more virulence genes than ST188-t189, and the prevalence of ST239 clone decreased with the proportion of MRSA among all *S. aureus* isolates.

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Introduction

Staphylococcus aureus (*S. aureus*) is an important pathogen causing a great number of infectious diseases. Recently, with the development of organ transplantation, bone marrow transplantation, and invasive diagnostic tests, bloodstream infections such as bacteremia and septicemia are on the rise. *S. aureus* is one of the most common human pathogen causing bloodstream infection, which can lead to both community- and hospital-acquired bacteremia. The mortality of patients with *S. aureus* bacteremia was reported to be about 30%,¹ which was closely associated with methicillin-resistant *S. aureus* (MRSA) infections.² Compared to patients with methicillin-sensitive *S. aureus* (MSSA) bacteremia, the mortality MRSA bacteremia increases by 40%.

It was observed that the molecular characteristics of *S. aureus* have regional differences. Sequence type 239 (ST239) were found to be the most prevalent in many Asian countries including China,^{3,4} but ST36 and ST30 were the most frequent in the United Kingdom.⁵ Researches about molecular characteristics and virulence gene profiles of bloodborne *S. aureus* isolates began late in China. A study carried out in 2012 which revealed that ST239-t030 and ST188-t189 were the most prevalent among the MRSA isolates, whereas ST7-t091 was the most prevalent among the MSSA isolates followed by ST188-t189 and ST630-t377.⁶ Subsequent studies showed that ST239-t030 and ST239-t037 were the most common in MRSA isolates, whereas ST7-t091 and ST188-t189 were the most prevalent in MSSA isolates.⁷⁻⁹ MRSA isolates carry more virulence encoding genes than MSSA isolates according to studies indicating that MRSA isolates have different molecular characteristics and virulence gene profiles compared to MSSA isolates.^{6,8,9}

Recently, with the increasing of awareness of infections control, the proportion of MRSA among all *S. aureus* isolates decreases significantly. China Antimicrobial Surveillance Network (CHINET) showed that the ratio of MRSA was 69.0% in 2005 and 35.3% in 2017.¹⁰ Because MRSA isolates differ from MSSA isolates in molecular characteristics and virulence genes profile, it is reasonable to speculate that the molecular characteristics and virulence genes profile of *S. aureus* isolates change as the proportion of MRSA among all *S. aureus* isolates decreases. The aims of this study were to investigate the molecular characteristics and virulence genes profile of bloodborne *S. aureus*, and analyze the correlation between the prevalence of the common STs and the proportion of MRSA among all *S. aureus* isolates. We also compared the differences of molecular characteristics and virulence genes profile between MRSA and MSSA isolates.

Material and methods

S. aureus isolation, identification and collection

A total of 112 non-duplicate *S. aureus* isolates were collected from eight hospitals in four cities from July 2017 to February 2018 (see Table 1). Isolates were identified as *S. aureus* using conventional microbiological methods including Gram staining, catalase and coagulase tests. They were further identified by Matrix-Assisted Laser Desorption Ionization Time of Flight

Table 1 – The distribution of 112 *S. aureus* isolates.

Hospital	No. of isolates	No. of MRSA isolates (% MRSA)
U	14	3 (21.4%)
Z	28	12 (42.9%)
P	11	3 (27.3%)
T	9	7 (77.8%)
J	12	0 (0.0%)
JC	4	2 (50.0%)
B	11	2 (18.2%)
H	23	3 (13.0%)
Total	112	32 (28.6%)

U, Union Hospital Affiliated to Tongji Medical College of Huazhong University of Science and Technology; Z, Zhongnan Hospital of Wuhan University; P, Puai Hospital Affiliated to Tongji Medical College of Huazhong University of Science and Technology; T, Tongji Hospital Affiliated to Tongji Medical College of Huazhong University of Science and Technology; J, the First People's Hospital of Jingzhou; JC, Jingzhou Central Hospital; B, Beijing Tsinghua Changgung Hospital Affiliated to Tsinghua University; H, Hainan General Hospital.

Mass Spectrometry (MALDI-TOF MS) System (BD, USA). An antimicrobial susceptibility test was first carried out using a VITEK 2 Compact system and VITEK 2 AST-GP67 Test Kit (bioMérieux, Inc., Durham, NC, USA). Methicillin resistance was confirmed using a cefoxitin disk (30 µg, Oxoid) and oxacillin disk (1 µg, Oxoid,) in Zhongnan Hospital, Wuhan university. All the isolates were stored at –80 °C for further experiments. The Ethics Committee of Zhongnan Hospital, Wuhan University approved this study. Because this was a retrospective study and all patients were anonymized, informed consent was waived.

S. aureus chromosomal DNA extraction

All *S. aureus* isolates were grown on blood agar at 37 °C overnight, then a single colony was transferred into 5 mL Tryptic Soy Broth (TSB) medium to culture for 16 h at a rotation speed of 200 rpm/min. *S. aureus* pellets were used to extract chromosomal DNA with TIANamp Bacteria DNA kit (Tiangen, China) supplemented with 1 mg/mL lysostaphin (Sigma, China) according to the manufacturer's instructions. All the extracted chromosomal DNAs were stored at –20 °C for further tests.

Staphylococcal protein A (*spa*) typing

The polymorphic X region of *spa* consists of a variable number of 24 bp repeat units allowing isolates to be distinguished from one another. In the present study, the polymorphic X region of *spa* was amplified from the extracted chromosomal DNAs using primers *spa*-1113f and *spa*-1514r¹¹ (see Table 2). The PCR mixture contained 2 µL *spa*-1113f (10 µM), 2 µL *spa*-1514r (10 µM), 1 µL chromosomal DNA template, 25 µL 2×Taq Master Mix (Tiangen, China), and 20 µL double distilled water. The PCR conditions were as follow: initial denaturation at 94 °C for 5 min, followed by 30 cycles of denaturation at 94 °C for 45 s, annealing at 53 °C for 60 s and extension at 72 °C for 90 s, and a final extension at 72 °C for 10 min. The amplicons

Table 2 – Primers used in this study and the PCR conditions for amplifying virulence genes.

Gene	Primer	Nucleotide sequence (5'–3')	Amplicon size (bp)	Annealing temperature	Extension time
<i>pvl</i>	PVL-F	ATCATTAGGTAAAATGTCTGGACATGATCCA	433	56 °C	30 s
	PVL-R	GCATCAASTGTATTGGATAGCAAAAAGC			
<i>fnbA</i>	FNBA-F	GTGAAGTTTTAGAAGGTGGAAAGATTAG	643	57 °C	40 s
	FNBA-R	GCTCTGTAAAGACCATTTTTCTTCCAC			
<i>fnbB</i>	FNBB-F	GTAACAGCTAATGGTCGAATTGATACT	524	57 °C	35 s
	FNBB-R	CAAGTTCGATAGGAGTACTATGTTC			
<i>hla</i>	HLA-F	CTGATTACTATCCAAGAAATTCGATTG	209	58 °C	15 s
	HLA-R	CTTTCAGCCTACTTTTTTATCAGT			
<i>hly</i>	HLB-F	GTGCCTTACTGACAATAGTGC	309	58 °C	20 s
	HLB-R	GTTGATGAGTAGCTACCTTCAGT			
<i>sea</i>	SEA-F	GAAAAAAGTCTGAATTGCAGGGAACA	560	55 °C	45 s
	SEA-R	CAAATAAATCGTAATTAACCGAAGGTTC			
<i>seb</i>	SEB-F	ATTCTATTAAGGACACTAAGTTAGGGA	404	57 °C	25 s
	SEB-R	ATCCCGTTTCATAAGGCGAGT			
<i>sec</i>	SEC-F	GTAAGTTACAGGTGGCAAACTTG	297	57 °C	20 s
	SEC-R	CATATCATACCAAAAAGTATTGCCGT			
<i>eta</i>	ETA-F	CGCTGCGGACATTCCTACATGG	676	57 °C	45 s
	ETA-R	TACATGCCCGCCACTTGCTTGT			
<i>etb</i>	ETB-F	CAGATAAAGAGCTTTATACACACATTAC	612	56 °C	45 s
	ETB-R	AGTGAACCTTATCTTTCTATTGAAAAACTC			
<i>tst</i>	TST-F	TTCACTATTGTAAAAGTGCAGACCCACT	180	57 °C	15 s
	TST-R	TACTAATGAATTTTTTATCGTAAGCCCTT			
<i>clfa</i>	CLFA-F	ATTGGCGTGGCTTCAGTGCT	292	57 °C	30 s
	CLFA-R	CGTTTCTCCGTAGTTGCATTG			
<i>spa</i>	Spa-1113f	TAAAGACGATCCTTCGGTGAGC	–	–	–
	Spa-1514r	CAGCAGTAGTGCCGTTTGCTT			

were sequenced (Tianyihuiyuan, China) and analyzed using the Ridom web server (<http://spaserver.ridom.de>).

Multilocus sequence typing (MLST)

MLST was carried out as described previously.¹² Seven *S. aureus* housekeeping genes (i.e. *arcC*, *aroE*, *glpF*, *gmk*, *pta*, *tpi* and *yqil*) were amplified by seven PCR assays followed by sequencing. The sequences of amplicons were compared to the known alleles stored in the MLST database (<http://saureus.mlst.net>) to identify the sequence type. Clustering of related STs, which were defined as clonal complexes (CCs), was determined using eBURST.

Detection of virulence genes

Several PCR assays were used to detect the following 12 staphylococcal virulence genes including the *pvl* genes (*lukF/S-PV*), the staphylococcal enterotoxin genes (*sea*, *seb*, *sec*), the exfoliative toxin genes (*eta*, *etb*), the hemolysin genes (*hla*, *hly*), the adhesion factor genes (*fnbA*, *fnbB*, *clfa*), and the toxic shock syndrome toxin (*tst*). All primers are listed in Table 2.^{13–15} The PCR mixture contained 2 µL of a couple of primers (10 µM), 1 µL DNA template, 12.5 µL Taq Master Mix (2×), and double distilled water (9.5 µL). The PCR conditions were as follow: initial denaturation at 94 °C for 5 min, followed by 30 cycles of denaturation at 94 °C for 30s, 30s with the respective annealing temperature, extension at 72 °C for respective extension time, and a final extension at 72 °C for 10 min. The annealing temperatures and extension times for different primers are shown in Table 2. All PCR products were analyzed by electrophoresis with 2% agarose gels.

Statistical analysis

Statistical analyses were performed using SPSS Statistics 24.0 for Windows. The chi-square test was used to compare the distributions of virulence genes between MRSA and MSSA isolates, ST5-t2460 and ST188-t189. The correlations between the prevalence of ST239, ST5 clones and the proportion of MRSA among all *S. aureus* isolates were analyzed using Spearman rank correlation. $p < 0.05$ was considered to be statistically significant.

Results and discussion

Molecular typing of *S. aureus* isolates.

The distribution of MLST and *spa* types of bloodborne *S. aureus* isolates is presented in Table 3. Among the 112 *S. aureus* isolates, 25 STs were identified. ST5 was the most predominant (21.4%), followed by ST188 (12.5%), ST59 (8.9%), ST398 (8.0%), ST1 (6.3%), ST630 (4.5%), and ST121 (4.5%). Other STs accounted for 33.9% of the 112 *S. aureus* isolates. In addition, one isolate could not be identified for ST. 95.8% of the ST5 isolates were collected from Wuhan, whereas 57.1% of the ST188 isolates were isolated from Haikou, which indicates that the distribution of STs of *S. aureus* have regional differences. Sixteen clone complexes (CCs) and three singletons were identified by eBURST (Fig. 1), CC5 (22.3%) was the most prevalent clonal complex, followed by CC8 (10.7%) and CC59 (9.8%). Thirty-two (28.6%) *S. aureus* isolates were MRSA in this study. The most major epidemic ST in MRSA isolates was ST5 (50.0%), followed by ST59 (15.6%), ST239 (12.5%), and ST45 (9.4%). In contrast, the most prevalent ST in MSSA isolates was

Table 3 – Molecular characteristics of *S. aureus* isolates from patients with bloodstream infection.

Clone complex (n)	MLST (n)	Spa (n)
CC5(25)	ST5(24)	t2460(16)
		t002(4)
		t548(1)
		t640(1)
		t4336(1)
		t5353(1)
CC188(14)	ST965(1)	t062(1)
	ST188(14)	t189(12)
		t8807(1)
		t4950(1)
CC8(12)	ST8(3)	t9101(2)
		t3641(1)
	ST239(4)	t030(4)
		ST630(5)
CC59(11)	ST59(10)	
		t13954(1)
CC398(9)	ST338(1)	t437(1)
	ST398(9)	t011(3)
		t034(3)
		t571(2)
		t1793(1)
CC1(7)	ST1(7)	t127(6)
CC121(5)	ST121(5)	t2246(1)
		t162(1)
		t269(1)
		t2086(1)
		t2091(1)
CC30(4)	ST30(4)	t1425(1)
		t012(2)
		t2868(1)
		NT(1)
CC15(4)	ST15(4)	t094(2)
		t085(1)
		t346(1)
		t116(2)
CC45(4)	ST45(4)	t550(1)
		NT(1)
		t796(1)
		t091(1)
CC7(3)	ST7(2)	t091(1)
		ST943(1)
		ST22(1)
		ST271(1)
CC22(2)	ST22(1)	t309(1)
CC72(2)	ST72(2)	t5763(1)
		t148(2)
CC88(2)	ST88(2)	t1376(1)
		t15796(1)
CC25(2)	ST25(2)	t227(1)
		t078(1)
CC20(2)	ST1281(2)	t164(2)
CG672(1)	ST672(1)	t3841(1)
CC6(1)	ST6(1)	t701(1)
CC2983(1)	ST2983(1)	t377(1)
NT(1)	NT(1)	t091(1)

NT, non-typeable; n, number of isolates in each type. The *spa* typing was based on variations of the repeat units, amplification and sequencing of the X region were performed as described previously by Strommenger et al.¹¹ MLST of *S. aureus* isolates was performed using detection of the 7 housekeeping genes of *S. aureus* as described previously.¹²

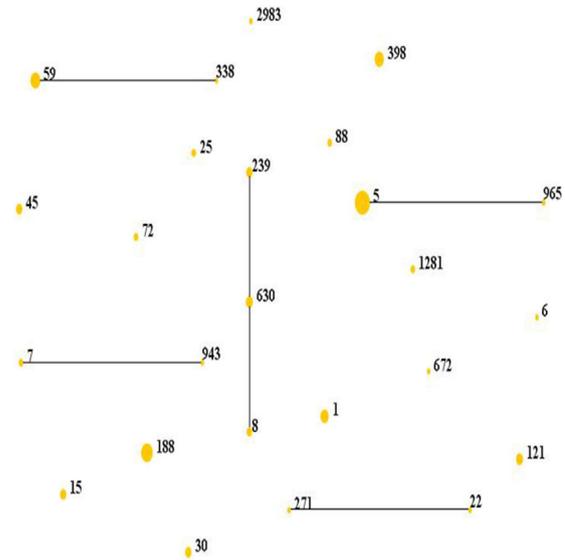


Fig. 1 – Distribution of STs in the clonal complexes. The diagram generated by eBURST with the default group definition based on the MLST data of this study, representing the relationships of 111 *S. aureus* isolates. Each number implies an MLST ST, STs that are linked by a line belong to the same cluster and the dot area indicates the prevalence of the ST in the MLST data of this study.

ST188 (17.5%) followed by ST398 (10.0%) and ST5 (10.0%). In last ten years in China, five studies about molecular characteristics of *S. aureus* isolates causing bloodstream infection were carried out.^{6,8,9,16,17} ST239 was found to be the most prevalent clone except in the studies by Yu and Li where ST239 was the most common clone among the MRSA isolates, suggesting that the prevalence of ST239 clone may be closely associated with MRSA infections. Twelve previous studies about molecular characteristic profiles of *S. aureus* isolates in China^{6,8,9,16-24} and our present study analyzed the correlation between the proportion of MRSA among all *S. aureus* isolates and the prevalence of ST239 and ST5 clone. All *S. aureus* isolates both MRSA and MSSA in the above studies were collected from patients rather than volunteers. The information of these *S. aureus* isolates is listed in Table 4. The rates of MRSA to total *S. aureus* isolates and the prevalence of ST239 and ST5 clone are shown in Fig. 2. Spearman rank correlation analysis showed that the proportion of MRSA among all *S. aureus* isolates correlated closely with the prevalence of ST239 clone ($r=0.8$, $p<0.001$) rather than ST5 clone ($r=0.2$, $p=0.5$), and the prevalence of ST239 clone decreased as the proportion of MRSA among all *S. aureus* isolates decreased, indicating that molecular characteristics of *S. aureus* isolates change as the proportion of MRSA among all *S. aureus* isolates fall. These studies also indicated that the implementation of infection control contributed to preventing the spread of ST239 *S. aureus* isolates.

Among the 112 *S. aureus* isolates, 48 *spa* types were found. The most prevalent *spa* type was t2460 (14.3%) followed by

Table 4 – Description of the *S. aureus* isolates included in 12 previous studies and our present study.

Number of studies	Source	Period	Participating cities	Participating hospitals	MRSA (%)	References
1	Various sources ^c	2014–2015	Nanchang	1	79.2	Chen et al., 2018
2	Various sources ^a	2005–2010	Shanghai	1	72.8	Song et al., 2013
3	Various sources ^d	2011	Beijing	1	70.2	Xiao et al., 2016
4	Blood	2009–2011	Shanghai	4	57.4	Chen et al., 2013
5	Skin and soft tissue infections	2002–2008	Wenzhou	1	54.1	Yao et al., 2010
6	Blood	2010–2011	Beijing, Tianjin, Shenyang, Jinan, Wuhan, Changsha, Hangzhou, Shanghai, Nanjing, Guangzhou, Xi'an and Chongqing,	16	47.5	He et al., 2013
7	Blood	2012–2016	Nanchang	1	46.4	Liu et al., 2018
8	Surgical site infections	2011–2012	Shanghai	1	43.9	Gu et al., 2015
9	Various sources ^b	2011–2012	Wuhan, Zhengzhou and Changsha	6	41.8	Liu et al., 2015
10	Blood	2004–2010	Wenzhou, Lishui, Shaoxing and Taizhou	4	41.6	Yu et al., 2012
11	Blood	2013 and 2016	Beijing, Changsha, Chongqing, Guangzhou, Hangzhou, Jinan, Nanjing, Shanghai, Shenyang, Tianjin, Wuhan and Xi'an	22	36.5	Li et al., 2018
12	Blood	2017–2018	Wuhan, Jingzhou, Beijing and Haikou	8	28.6	Our present study
13	Nasal cavity	2015	Shanghai	1	27.3	Chen et al., 2016

^a Abscesses or wounds, sputum, blood and other fluids (abdominal fluid, cerebrospinal fluid and peritoneal dialysis fluid).

^b Blood, cerebrospinal fluid, ascites, pleural effusion, and synovial fluid.

^c Skin and soft tissue infections, respiratory tract, blood, unknown.

^d Pneumonia, soft tissue infections, urinary tract infections, bloodstream infections, joint infections and gallbladder infections.

t189 (10.7%), t437 (8.9%), t377 (5.4%), t127 (5.4%), t002 (3.6%), and t030 (3.6%). In addition, two isolates could not be identified for *spa*. All the t2460 *S. aureus* isolates were collected from Wuhan except one isolate from Beijing, whereas the t189 *S. aureus* isolates were collected in Haikou (58.3%), Wuhan (25.0%), Jingzhou (8.3%) and Beijing (8.3%). In 2007, t2460 clone was first found in South Korea,²⁵ whereas in China it was not be found in large numbers until May 2015.²⁶ These data may supported the view of the temporary outbreak of t2460

in Wuhan. To date, t091 and t030 were the most common *spa* types in bloodstream infection in China,^{6–9} whereas t437 and t127 were found to be the most common *spa* types in other infections.^{27–29}

Virulence gene profiles

The pathogenicity of *S. aureus* is closely related to presence of various virulence genes.³⁰ The frequencies of virulence

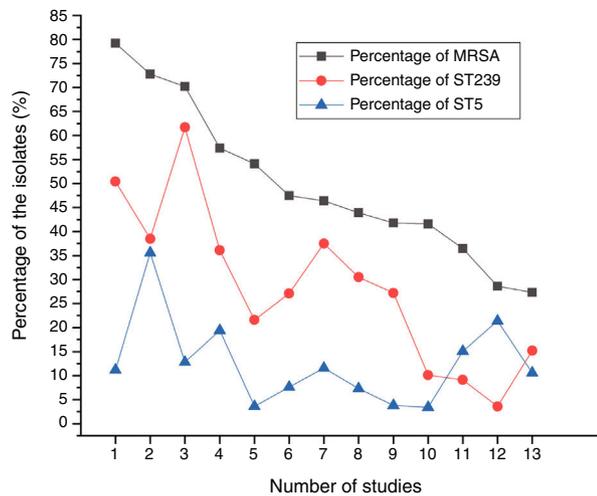


Fig. 2 – The proportions of MRSA among all *S. aureus* isolates and the prevalence of ST239 or ST5 clone.

encoding genes identified in the 112 isolates are listed in Table 5. Our data showed that all *S. aureus* isolates carried at least two virulence genes and almost all the isolates contained *clfa* (100%, 112/112) and *hla* (99.1%, 111/112) genes. These findings are comparable to those reported in many hospitals of south of China,^{31,32} which indicated that *clfa* and *hla* were widely present and may play a pivotal role in the pathogenicity of *S. aureus*. Enterotoxins, including *sea*, *seb* and *sec* were found to cause food poisoning primarily. In our study, the most common enterotoxin gene was *seb* (66.1%), which is significantly different from those reported in foodborne *S. aureus*, where the most common enterotoxin gene is *sea*.^{33,34} Our study showed that 55.4% (62/112) isolates harbored six or more virulence genes, of which two isolates contained nine virulence genes, 15 isolates harbored eight virulence genes, 15 isolates carried eight virulence genes, 18 isolates had seven virulence genes, 27 isolates harbored six virulence genes. Furthermore, six or more virulence genes were present in 81.3% of MRSA isolates and 45% of MSSA isolates and MRSA isolates harbored more virulence genes than the MSSA isolates ($\chi^2 = 12.154$, $p < 0.01$).

The above results suggested that *S. aureus* isolates with different genetic background have different ability to acquire mobile genetic elements such as plasmids, phages and pathogenicity islands.

Molecular characteristics of the major clones ST5-t2460 and ST188-t189

MLST typing and *spa* typing were performed to analyze the molecular characteristics of *S. aureus* isolates. There was a strong association observed between ST and *spa* types: the ST5 type was primarily associated with t2460 (66.7%, 16/24), the ST188 type with t189 (85.7%, 12/14), the ST59 type with t437 (90%, 9/10), the ST1 type with t127 (85.7%, 6/7) and the ST630 type with t377 (100%, 5/5). ST5-t2460 (16/112, 14.3%) were found to be the most common in this study, followed by ST188-t189 (12/112, 10.7%). Molecular characteristics of ST5-t2460 and ST188-t189 are summarized in Table 6. 93.8% of ST5-t2460 were collected from four hospitals in Wuhan, whereas the distribution of ST188-t189 was more dispersed including in Haikou (58.3%), Wuhan (25%), Beijing (8.3%) and Jingzhou (8.3%). In 2007, ST5-t2460 became the main type of MRSA bloodstream infections in Korea which had never been seen before.²⁵ In China, extremely few cases of ST5-t2460 were reported in previous studies.^{16,35} Therefore, the prevalence of ST5-t2460 in our research may be consequence of a temporary outbreak of ST5-t2460 in Wuhan. In our present study, all the ST5-t2460 were MRSA, whereas all the ST188-t189 were MSSA. The prevalence of strains with six or more virulence genes was significantly higher in ST5-t2460 strains (93.8%) compared with ST188-t189 strains (8.3%) ($p < 0.001$). In addition, of the tested 12 virulence genes, *pvl*, *fnbA*, *hlyB*, *sec*, and *tst* were found to be more frequent in ST5-t2460 than in ST188-t189. These results indicated that ST5-t2460 acquire *mecA* and virulence genes easily, which supported the above conclusion that *S. aureus* with different genetic background have different ability to acquire mobile genetic elements. In addition to genetic background, ST5-t2460 often lead to persistent and recurrent MRSA bloodstream infections,³⁶ which also contribute to promoting *S. aureus* isolates to acquire virulence genes easily.

Table 5 – The frequencies of virulence encoding genes among *S. aureus*, MRSA and MSSA isolates in bloodstream infection.

Virulence genes	<i>S. aureus</i> (N = 112) n(%)	MRSA(N = 32) n(%)	MSSA(N = 80) n(%)	p-Value ^a
<i>pvl</i>	49 (43.8)	17 (53.1)	32 (40)	0.206
<i>fnbA</i>	70 (62.5)	25 (78.1)	45 (56.3)	0.031
<i>fnbB</i>	33 (29.5)	4 (12.5)	29 (36.3)	0.013
<i>hla</i>	111 (99.1)	32 (100)	79 (98.8)	1.000
<i>hlyB</i>	59 (52.7)	25 (78.1)	34 (42.5)	0.001
<i>sea</i>	19 (17.0)	7 (21.9)	12 (15)	0.629
<i>seb</i>	74 (66.1)	21 (65.6)	53 (66.3)	0.950
<i>sec</i>	47 (42.0)	22 (68.8)	25 (31.3)	<0.001
<i>eta</i>	17 (15.2)	5 (15.6)	12 (15)	1.000
<i>etb</i>	2 (1.8)	0 (0)	2 (2.5)	1.000
<i>tst</i>	36 (32.1)	18 (56.3)	17 (21.3)	<0.001
<i>clfa</i>	112 (100)	32 (100)	80 (100)	–

^a The positive rates of virulence genes in MRSA isolates were compared with those in MSSA isolates.

Table 6 – The frequencies of virulence encoding genes among *S. aureus*, ST5-t2460 *S. aureus* and ST188-t189 *S. aureus* isolates in bloodstream infection.

	<i>pvl</i>	<i>fnbA</i>	<i>fnbB</i>	<i>hla</i>	<i>hly</i>	<i>sea</i>	<i>seb</i>	<i>sec</i>	<i>eta</i>	<i>etb</i>	<i>tst</i>	<i>clfA</i>	MRSA
ST5-t2460 (N = 16) n(%)	12 (15)	14 (87.5)	1 (6.3)	16 (100)	16 (100)	0 (0)	10 (62.5)	16 (100)	0 (0)	0 (0)	16 (100)	16 (100)	16 (100)
ST188-t189 (N = 12) n(%)	3 (25)	6 (50)	1 (8.3)	12 (100)	4 (33.3)	0 (0)	9 (75)	0 (0)	0 (0)	0 (0)	1 (8.3)	12 (100)	0 (0)
<i>p</i> value ^a	0.020	0.044	1.000	–	<0.001	–	0.678	<0.001	–	–	<0.001	–	<0.001

^a The positive rates of virulence genes and MRSA among ST5-t2460 strains were compared with those among ST188-t189 strains.

In conclusion, ST5-t2460 was the most common clone in *S. aureus* causing bloodstream infection, followed by ST188-t189, which had never been reported in China before. All the ST5-t2460 were MRSA, whereas all the ST188-t189 were MSSA. Moreover, ST5-t2460 harbored more virulence genes than ST188-t189, which indicates that ST5-t2460 acquire *mecA* and virulence genes more easily than ST188-t189. In addition, the prevalence of ST239 clone decreased with the decrease of the proportion of MRSA among all *S. aureus* isolates.

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Conflicts of interest

The authors declare no conflicts of interest.

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REFERENCES

- De Rosa FG, Corcione S, Motta I, et al. Risk factors for mortality in patients with *Staphylococcus aureus* bloodstream infection. *J Chemother*. 2016;28:187–90.
- Shurland S, Zhan M, Bradham DD, Roghmann MC. Comparison of mortality risk associated with bacteremia due to methicillin-resistant and methicillin-susceptible *Staphylococcus aureus*. *Infect Control Hosp Epidemiol*. 2007;28:273–9.
- Cha HY, Moon DC, Choi CH, et al. Prevalence of the ST239 clone of methicillin-resistant *Staphylococcus aureus* and differences in antimicrobial susceptibilities of ST239 and ST5 clones identified in a Korean hospital. *J Clin Microbiol*. 2005;43:3610–4.
- Zhang W, Shen X, Zhang H, et al. Molecular epidemiological analysis of methicillin-resistant *Staphylococcus aureus* isolates from Chinese pediatric patients. *Eur J Clin Microbiol Infect Dis*. 2009;28:861–4.
- Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol*. 2000;38:1008–15.
- Yu F, Li T, Huang X, et al. Virulence gene profiling and molecular characterization of hospital-acquired *Staphylococcus aureus* isolates associated with bloodstream infection. *Diagn Microbiol Infect Dis*. 2012;74:363–8.
- Wang WY, Chiueh TS, Sun JR, Tsao SM, Lu JJ. Molecular typing and phenotype characterization of methicillin-resistant *Staphylococcus aureus* isolates from blood in Taiwan. *PLoS ONE*. 2012;7:e30394.
- Chen X, Wang WK, Han LZ, et al. Epidemiological and genetic diversity of *Staphylococcus aureus* causing bloodstream infection in Shanghai, 2009–2011. *PLoS ONE*. 2013;8:e72811.
- He W, Chen H, Zhao C, et al. Population structure and characterisation of *Staphylococcus aureus* from bacteraemia at multiple hospitals in China: association between antimicrobial resistance, toxin genes and genotypes. *Int J Antimicrob Agents*. 2013;42:211–9.
- Hu F, Guo Y, Zhu D, et al. Antimicrobial resistance profile of clinical isolates in hospitals across China: report from the CHINET Surveillance Program, 2017. *Chin J Infect Chemother*. 2018;18:241–51.
- Strommenger B, Kettlitz C, Weniger T, Harmsen D, Friedrich AW, Witte W. Assignment of *Staphylococcus* isolates to groups by *spa* typing SmaI macrorestriction analysis, and multilocus sequence typing. *J Clin Microbiol*. 2006;44:2533–40.
- Saunders NA, Holmes A. Multilocus sequence typing (MLST) of *Staphylococcus aureus*. *Methods Mol Biol*. 2007;391:71–85.
- Jarraud S, Mougel C, Thioulouse J, et al. Relationships between *Staphylococcus aureus* genetic background, virulence factors, *agr* groups (alleles), and human disease. *Infect Immun*. 2002;70:631–41.
- Tristan A, Ying L, Bes M, Etienne J, Vandenesch F, Lina G. Use of multiplex PCR to identify *Staphylococcus aureus* adhesins involved in human hematogenous infections. *J Clin Microbiol*. 2003;41:4465–7.
- Campbell SJ, Deshmukh HS, Nelson CL, et al. Genotypic characteristics of *Staphylococcus aureus* isolates from a multinational trial of complicated skin and skin structure infections. *J Clin Microbiol*. 2008;46:678–84.
- Li S, Sun S, Yang C, et al. The changing pattern of population structure of *Staphylococcus aureus* from bacteremia in China from 2013 to 2016: ST239-030-MRSA replaced by ST59-t437. *Front Microbiol*. 2018;9:332.
- Liu Y, Du FL, Liu PP, et al. Molecular epidemiology and virulence features of *Staphylococcus aureus* bloodstream isolates in a regional burn center in China, 2012–2016. *Microb Drug Resist*. 2018.
- Yao D, Yu FY, Qin ZQ, et al. Molecular characterization of *Staphylococcus aureus* isolates causing skin and soft tissue infections (SSTIs). *BMC Infect Dis*. 2010;10:133.
- Gu F-F, Han L-Z, Chen X, et al. Molecular characterization of *Staphylococcus aureus* from surgical site infections in orthopedic patients in an orthopedic trauma clinical medical center in Shanghai. *Surg Infect (Larchmt)*. 2015;16:97–104.

20. Song Y, Du X, Li T, Zhu Y, Li M. Phenotypic and molecular characterization of *Staphylococcus aureus* recovered from different clinical specimens of inpatients at a teaching hospital in Shanghai between 2005 and 2010. *J Med Microbiol.* 2013;62:274–82.
21. Liu C, Chen ZJ, Sun Z, et al. Molecular characteristics and virulence factors in methicillin-susceptible, resistant, and heterogeneous vancomycin-intermediate *Staphylococcus aureus* from central-southern China. *J Microbiol Immunol Infect.* 2015;48:490–6.
22. Chen X, Sun K, Dong D, Luo Q, Peng Y, Chen F. Antimicrobial resistance and molecular characteristics of nasal *Staphylococcus aureus* isolates from newly admitted inpatients. *Ann Lab Med.* 2016;36:250.
23. Chen K, Lin S, Li P, et al. Characterization of *Staphylococcus aureus* isolated from patients with burns in a regional burn center, Southeastern China. *BMC Infect Dis.* 2018;18:51.
24. Xiao M, Zhao R, Zhang Q, et al. Genotypic Diversity of *Staphylococcus aureus* α -hemolysin gene (hla) and its association with clonal background: implications for vaccine development. *PLOS ONE.* 2016;11:e0149112.
25. Kim T, Yi J, Hong KH, Park JS, Kim EC. Distribution of virulence genes in spa types of methicillin-resistant *Staphylococcus aureus* isolated from patients in intensive care units. *Korean J Lab Med.* 2011;31:30–6.
26. Li Y, Zhao R, Zhang X, et al. Prevalence of enterotoxin genes and spa genotypes of methicillin-resistant *Staphylococcus aureus* from a tertiary care hospital in China. *J Clin Diagn Res.* 2015;9:DC11–4.
27. Li T, Yu X, Xie J, et al. Carriage of virulence factors and molecular characteristics of *Staphylococcus aureus* isolates associated with bloodstream, and skin and soft tissue infections in children. *Epidemiol Infect.* 2013;141:2158–62.
28. Zhang J, Gu F-F, Zhao S-Y, et al. Prevalence and molecular epidemiology of *Staphylococcus aureus* among residents of seven nursing homes in Shanghai. *PLOS ONE.* 2015;10:e0137593.
29. Wang X, Li X, Liu W, Huang W, Fu Q, Li M. Molecular characteristic and virulence gene profiles of community-associated methicillin-resistant *Staphylococcus aureus* isolates from pediatric patients in Shanghai China. *Front Microbiol.* 2016;7:1818.
30. Yu F, Yang L, Pan J, et al. Prevalence of virulence genes among invasive and colonising *Staphylococcus aureus* isolates. *J Hosp Infect.* 2011;77:89–91.
31. Xie X, Dai X, Ni L, et al. Molecular epidemiology and virulence characteristics of *Staphylococcus aureus* nasal colonization in medical laboratory staff: comparison between microbiological and non-microbiological laboratories. *BMC Infect Dis.* 2018;18:122.
32. Jiang B, Yin S, You B, et al. Antimicrobial resistance and virulence genes profiling of methicillin-resistant *Staphylococcus aureus* isolates in a burn center: a 5-year study. *Microb Pathog.* 2018;114:176–9.
33. Argudin MA, Mendoza MC, Gonzalez-Hevia MA, Bances M, Guerra B, Rodicio MR. Genotypes, exotoxin gene content, and antimicrobial resistance of *Staphylococcus aureus* strains recovered from foods and food handlers. *Appl Environ Microbiol.* 2012;78:2930–5.
34. Sato'o Y, Omoe K, Naito I, et al. Molecular epidemiology and identification of a *Staphylococcus aureus* clone causing food poisoning outbreaks in Japan. *J Clin Microbiol.* 2014;52:2637–40.
35. Yu F, Liu Y, Lv J, et al. Antimicrobial susceptibility, virulence determinant carriage and molecular characteristics of *Staphylococcus aureus* isolates associated with skin and soft tissue infections. *Braz J Infect Dis.* 2015;19: 614–22.
36. Kim NH, Kang YM, Han WD, et al. Small-colony variants in persistent and recurrent *Staphylococcus aureus* bacteremia. *Microb Drug Resist.* 2016;22:538–44.