



The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Original article

Prevalence of high-risk human papillomavirus by cobas 4800 HPV test in urban Peru



Ricardo Iwasaki*, Felipe Galvez-Philpott, Javier Arias-Stella Jr., Javier Arias-Stella

Precisa Laboratorios Arias Stella, Lima, Peru

ARTICLE INFO

Article history:

Received 3 December 2013

Accepted 24 January 2014

Available online 15 May 2014

Keywords:

High-risk human papillomavirus

Prevalence

Cobas 4800 HPV test

ABSTRACT

Background: Molecular tests allow the detection of high-risk human papillomavirus in cervical samples, playing an important role in the prevention of cervical cancer.

Objectives: We performed a study to determine the prevalence of HPV 16, HPV 18 and other high-risk human papillomavirus (pool 12 genotypes) in Peruvian females from diverse urban areas using the cobas 4800 HPV test.

Methods: Routine cervical samples collected in our laboratory were analyzed by cobas 4800 HPV test.

Results: A total of 2247 samples from female patients aged 17–79 years were tested. high-risk human papillomavirus was positive in 775 (34.49%) samples. Of these, 641 (82.71%) were single infections and 134 (17.29%) were multiple infections. The positivity rates for HPV 16, HPV 18, and other high-risk human papillomavirus were 10.77%, 2.0%, and 28.08%, respectively. In multiple high-risk human papillomavirus infections, the concomitance of HPV 16 and other high-risk human papillomavirus was more prevalent (13.42%).

Conclusion: Our study showed high prevalence of high-risk human papillomavirus in urban Peru, mainly among young women. In both single and multiple infections other high-risk human papillomavirus were more prevalent than HPV 16 and HPV 18, which might influence vaccine impact in our country. Furthermore, the cobas 4800 HPV test may be considered a useful tool for HPV molecular diagnosis.

© 2014 Elsevier Editora Ltda. Este é um artigo Open Access sob a licença de [CC BY-NC-ND](http://creativecommons.org/licenses/by-nc-nd/4.0/)

Introduction

Cervical cancer is the second most common cause of cancer in women¹ and it is estimated that each year approximately 493,000 new cases are diagnosed and 274,000 women die from cervical cancer worldwide.² Human Papillomavirus (HPV) has been recognized as an important cause of cervical cancer³ and

is implicated in 99.7% of cervical squamous cell cancer cases in the world.⁴ More than 40 HPV genotypes have been detected in the anogenital mucosa and are usually transmitted through sexual activity.⁵ The HPV genotypes 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68 are associated with cervical cancer and have been classified as high-risk (HR) group.^{6,7}

A global meta-analysis showed that the prevalence of HPV in 157,879 women with normal cytology was 10.4%. The

* Corresponding author at: Av. Gregorio Escobedo, 612, Jesús María, Lima 11, Peru.

E-mail address: biomolecular@ariasstella.com (R. Iwasaki).

<http://dx.doi.org/10.1016/j.bjid.2014.01.010>

1413-8670/© 2014 Elsevier Editora Ltda. Este é um artigo Open Access sob a licença de [CC BY-NC-ND](http://creativecommons.org/licenses/by-nc-nd/4.0/)

prevalence estimates by region were Africa 22.1%, Central America and Mexico 20.4%, Northern America 11.3%, Europe 8.1%, and Asia 8.0%.⁸ Of all high-risk human papillomavirus (HR-HPV) genotypes, HPV 16 has been regarded as the most prevalent genotypes.⁹

Few studies have examined the epidemiology of HPV in Peru. In the Department of San Martin, 5435 women were tested for HPV by Hybrid Capture II (Qiagen, Gaithersburg, MD); the prevalence of HPV was estimated to be 12.6%.¹⁰ On the other hand, the overall presence of HR-HPV was 33.6% in a study of 2208 women with abnormal cytology from different cities in Peru.¹¹ Also, an analysis performed on 384 cervical samples from six regions with the highest percentage of rural inhabitants in Peru showed the presence of HPV in 73 cases (19.01%) and the most common genotype was HPV 16, found in six (8.22%) cases.¹²

The DNA testing of HR-HPV genotypes in cervical samples has a sensitivity of up to 96.6% for detecting precancerous lesions.¹³ The cobas 4800 HPV Test (Roche Molecular Systems, Inc) is a qualitative *in vitro* test for the detection of HPV in patient specimens. The test utilizes amplification of target DNA by real time PCR and nucleic acid hybridization for the detection of 14 HR-HPV genotypes in a single analysis. The test specifically identifies genotypes 16 and 18 while simultaneously detecting the other HR genotypes (pool of 12 genotypes: 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68). The ATHENA (Addressing THE Need for Advanced HPV Diagnostics) study, involving more than 47,000 women in the United States, demonstrated that the cobas 4800 HPV Test is clinically validated for ASCUS triage.¹⁴

The present study aims to determine the prevalence of HPV 16, HPV 18 and other HR-HPV (pool of 12 genotypes) in cervical scrapes from women living in diverse urban areas in Peru by the cobas 4800 HPV Test.

Materials and methods

Study population

During the period from November 2011 to October 2013, routine cervical samples collected from women living in different urban areas across the country were received at the laboratory. Urban areas include the cities of Iquitos, Cajamarca, Piura, Chiclayo, Lima, Arequipa, Cuzco and Juliaca. On date of arrival, the samples were registered and labeled with a barcode for personal identification.

Cobas 4800 HPV test

The cobas 4800 HPV test is a qualitative assay for the presence of HPV 16, HPV 18 and a pool of 12 other HR-HPV genotypes (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68). This test requires 1 mL of liquid cytology sample. No pre-treatment of samples is required. The cobas 4800 HPV test was performed according to the recommendations of the manufacturer.¹⁵ Briefly, DNA was extracted, purified and prepared for PCR by the cobas x 480 instrument. Amplification and detection of HR-HPV DNA were undertaken on the cobas z 480 analyzer. Fluorescent TaqMan

probes were used for detection of the amplicons during PCR cycles.

Both positive and negative control specimens are included in each run. In addition, the test detects beta-globin DNA as an internal control of sufficient specimen cellularity. Specialized pipetting technology combined with AmpErase enzymes reduces cross contamination risk.

Analysis

HPV detection in cervical samples was considered as positive or negative. Prevalence of HPV including HPV 16, HPV 18 and other HR-HPV (pool of 12 genotypes) was reported. The frequencies of single and multiple HPV infections were also described. The results of the cobas testing were tabulated by age of the female patients.

Results

A total of 2247 cervical samples were analyzed. Patients were with the age 17–79 years (average 34.48 years). All cervical samples contained sufficient DNA to perform PCR, based on amplification of the human beta-globin gene. In the present study, HR-HPV was detected in 775 samples and the overall prevalence was 34.49% (Table 1). HPV 16 was detected in 242 samples, HPV 18 in 45 samples, and other HR-HPV (pool of 12 genotypes) in 631 samples, either as single or multiple infections combined (Table 2). The prevalence of other HR-HPV (28.08%) was higher than HPV 16 (10.77%) and HPV 18 (2.0%).

Of 775 HPV positive samples, 641 (82.71%) were single infections and 134 (17.29%) were multiple infections. Other HR-HPV (pool of 12 genotypes) were the most common in single (500/641) and in multiple infections (131/134). In single infections, HPV 16 was detected in 126 samples, whereas HPV 18 in 15. With respect to multiple HPV infections, HPV 16 and other HR-HPV were the most common (13.42%) coinfections, while HPV 16 and HPV 18 coinfections were the least common

Table 1 – Prevalence of HR-HPV among 2247 samples.

Result of cobas testing	Samples	Prevalence (%)
HR-HPV ^a	775	34.49
Negative	1472	65.51
Total	2247	100.00

^a HPV 16, 18 and other HR-HPV (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68).

Table 2 – Frequency of HPV 16, 18 and other HR-HPV.

Outcomes of cobas testing	Samples	Frequency (%) ^c
HPV 16 ^a	242	10.77
HPV 18 ^a	45	2.00
Other HR-HPV ^{a,b}	631	28.08

^a Single and multiple infections combined.

^b Genotypes 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68.

^c HPV positive in the total of samples.

Table 3 – Single or multiple infections in 775 HPV positive samples.

Genotype	Single infections		Relationship	Multiple infections	
	n	% ^b		n	% ^b
HPV 16	126	16.26	HPV 16/HPV 18	3	0.39
HPV 18	15	1.93	HPV 16/other HR-HPV ^a	104	13.42
Other HR-HPV ^a	500	64.52	HPV 18/other HR-HPV ^a	18	2.32
–	–	–	HPV16/HPV18/other HR-HPV ^a	9	1.16
Total	641	82.71	Total	134	17.29

^a Genotypes 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68.

^b Frequency: n among total HPV positive samples.

Table 4 – HR-HPV detection according to age.

Age (years)	HR-HPV ^a	Prevalence (%)
17–29	334	43.10
30–39	290	37.42
40–49	108	13.93
50–59	34	4.39
60–79	9	1.16
Total	775	100.00

^a HPV 16, 18 and other HR-HPV (31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68).

(0.39%). Also we observed the simultaneous presence of HPV 16, HPV 18 and other HR-HPV at least in nine samples (Table 3).

The prevalence of HR-HPV was highest in younger women and showed a decreasing trend with age (Table 4). The prevalence according to age was as follows: 43.1% for women under 29 years; 37.42% for those between 30 and 39 years; 13.94% in the age range 40–49; 4.39% between 50 and 59; and 1.16% among those aged 60–79.

Discussion

The high prevalence of HR-HPV (34.49%) in women from different urban areas of Peru is similar to that described in other countries such as Russia, 29.1%; Trinidad and Tobago, 35.4%; and East African countries, 33.6%.^{8,16} But it was higher in other parts of Latin America, such as Mexico, 14.5%; Costa Rica, 16.0%; Colombia, 20.5%; and Ecuador, 24.2%.^{17–20} However, it was lower than the rate found in Brazil (Rio de Janeiro).²¹ As it happens in most developing countries, some socio-economic and cultural factors may be related to this high HPV prevalence, especially early start of sexual activity; multiparity; lack of education; and the misconceptions and beliefs that constrain people from discussing diseases of the genital tract.²²

In Peru, previous studies found that the prevalence of HPV in the Department of San Martín was 12.6%¹⁰ and in some rural areas was 19.01%,¹² which were lower than the prevalence obtained in the present study. However, our data are consistent with those obtained by Li Ning et al.,¹¹ where the prevalence of HR-HPV in different cities of our country was 33.6%.

This is the first study about prevalence of HR-HPV using the cobas 4800 HPV Test in our country. As a part of our routine, cervical samples were received for HPV molecular

diagnosis. The test identifies the presence of HPV 16, HPV 18 and other HR-HPV (pool of 12 genotypes), which are involved with cervical cancer.²³ In both single and multiple infections, other HR-HPV were more common than HPV 16 and HPV 18, similar to a previous study in 5072 women from Denmark.²⁴ This is because infections by other HPV genotypes (not 16/18) are more frequent in women with normal cytology or low-grade CIN, although these genotypes only cause about one-third of cervical cancer.²⁵ Furthermore, our results are in line with studies indicating higher prevalence of HR-HPV in young women (17–29 years) and HPV infection decreasing with aging.^{24,26,27} However, it is important to identify which are the most common genotypes in the group of other HR-HPV. This would help to include the prevailing genotypes in the design of multivalent vaccines. It is also desirable to incorporate these genotypes in the next generation of automated molecular diagnostic systems.

Several studies assessed the use of the cobas 4800 HPV test.^{24,28–31} Those studies showed that the test have a high specificity and sensitivity, less cross contamination risk, clinical relevance, sufficient interlaboratory reproducibility, and less labor intensive. Additionally, this test fulfills all requirements as defined by the international guidelines³² when assessing a test for screening purposes.

In conclusion, our study showed a high prevalence of HR-HPV in females from diverse urban areas of Peru especially among young women. Both single and multiple HR-HPV infections underscore the presence of HR-HPV (pool of 12 genotypes) in excess of HPV 16 and HPV 18. This finding might influence vaccine impact in our country. Finally, the cobas 4800 HPV Test proved to be a useful tool for HPV molecular diagnosis.

Conflicts of interest

The authors declare no conflicts of interest.

REFERENCES

1. Pisani P, Bray F, Parkin DM. Estimates of the world-wide prevalence of cancer for 25 sites in the adult population. *Int J Cancer*. 2002;97:72–81.
2. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics. *CA Cancer J Clin*. 2005;55:74–108.

3. Burd EM. Human papillomavirus and cervical cancer. *Clin Microbiol Rev.* 2003;16:1-17.
4. Walboomers JM, Jacobs MV, Manos MM, et al. Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. *J Pathol.* 1999;189:12-9.
5. zur Hausen H. Papillomavirus infections—a major cause of human cancers. *Biochim Biophys Acta.* 1996;1288:55-78.
6. Bosch FX, Manos MM, Munoz N, et al. Prevalence of human papillomavirus in cervical cancer: a worldwide perspective. *J Natl Cancer Inst.* 1995;87:796-802.
7. Jacob MV, Snijders PJ, van den Brule AJ, Helmerhorst TJ, Meijer CJ, Walboomers JM. A general primer GP5+/GP6(+)-mediated PCR-enzyme immunoassay method for rapid detection of 14 high-risk and 6 low-risk human papillomavirus genotypes in cervical scrapings. *J Clin Microbiol.* 1997;35:791-5.
8. de Sanjose S, Diaz M, Castellsague X, et al. Worldwide prevalence and genotype distribution of cervical human papillomavirus DNA in women with normal cytology: a meta-analysis. *Lancet Infect Dis.* 2007;7:453-9.
9. Louvanto K, Rintala MA, Syrjanen KJ, Grenman SE, Syrjanen SM. Genotype-specific persistence of genital human papillomavirus (HPV) infections in women followed for 6 years in the Finnish Family HPV Study. *J Infect Dis.* 2010;202:436-44.
10. Almonte M, Ferreccio C, Winkler JL, et al. Cervical screening by visual inspection, VPH testing, liquid-based and conventional cytology in Amazonian Peru. *Int J Cancer.* 2007;121:796-802.
11. Li Ning ML, Bazan M, Arias-Stella Jr J. HPV DNA testing in a population with high prevalence of cervical squamous carcinoma: 5-year experience in urban Peru. Denver, CO: USCAP: United States & Canadian Academy of Pathology; 2008. p. 1-7.
12. Concha-M R, Arias-Stella Jr J, Quiñones D, Bazan M, Iwasaki R, Arias-Stella J. Investigación del ADN del virus del papiloma humano en el cuello uterino en población rural del Perú. *Patología Rev Latinoam.* 2012;50:266-71.
13. Mayrand MH, Duarte-Franco E, Rodrigues I, et al. Human papillomavirus DNA versus Papanicolaou screening tests for cervical cancer. *N Engl J Med.* 2007;357:1579-88.
14. Stoler MH, Wright Jr TC, Sharma A, Apple R, Gutekunst K, Wright TL. High-risk human papillomavirus testing in women with ASC-US cytology: results from the ATHENA HPV Study. *Am J Clin Pathol.* 2011;135:468-75.
15. Cobas 4800 HPV Test (package insert). Branchburg, NJ, USA: Roche Molecular Systems. Available: <http://e-labdoc.roche.com>
16. Bruni L, Diaz M, Castellsague M, Ferrer E, Bosch FX, de Sanjose S. Cervical human papillomavirus prevalence in 5 continents: meta-analysis of 1 million women with normal cytological findings. *J Infect Dis.* 2010;202:1789-99.
17. Lazcano-Ponce E, Herrero R, Munoz N, et al. Epidemiology of HPV infection among Mexican women with normal cervical cytology. *Int J Cancer.* 2001;91:412-20.
18. Herrero R, Hildesheim A, Bratti C, et al. Population-based study of human papillomavirus infection and cervical neoplasia in rural Costa Rica. *J Natl Cancer Inst.* 2000;92:464-74.
19. Farfan-Vargas YA, Garcia-Robayo DA, Arias-Murillo Y, Morales OL, Isaza M, Aristizabal-Gutierrez FA. Genotipificación del virus de papiloma humano en mujeres con hallazgo citológico de lesión escamosa intraepitelial de bajo grado (Isil) o de significado indeterminado (asc-us) en Bogotá, Colombia. *Rev Colomb Cienc Quím Farm.* 2010;39:42-54.
20. Brown CR, Leon ML, Munoz K, et al. Human papillomavirus infection and its association with cervical dysplasia in Ecuadorian women attending a private cancer screening clinic. *Braz J Med Biol Res.* 2009;42:629-36.
21. Carvalho MOO, Carestiato FN, Perdigo PH, et al. Human papillomavirus infection in Rio de Janeiro, Brazil: a retrospective study. *Braz J Infect Dis.* 2005;9:398-404.
22. Tabora N, Bakkers JM, Quint WG, et al. Human papillomavirus infection in Honduran women with normal cytology. *Cancer Causes Control.* 2009;20:1663-70.
23. Clifford GM, Smith JS, Plummer M, Munoz N, Franceschi S. Human papillomavirus types in invasive cervical cancer worldwide: a meta-analysis. *Br J Cancer.* 2003;88:63-73.
24. Preisler S, Rebolj M, Untermann A, et al. Prevalence of human papillomavirus in 5,072 consecutive cervical SurePath samples evaluated with the Roche cobas HPV real-time PCR assay. *PLOS ONE.* 2013;8:1-8.
25. Munoz N, Bosch FX, Castellsague X, et al. Against which human papillomavirus types shall we vaccinate and screen? The international perspective. *Int J Cancer.* 2004;111:278-85.
26. Dunne EF, Unger ER, Sternberg M, et al. Prevalence of HPV infection among females in the United States. *JAMA.* 2007;297:813-9.
27. Bedia D, Rotal S, Onan A, et al. Prevalence of human papillomavirus (HPV) and HPV-16 genotyping by real-time PCR in patients with several cervical pathologies. *Braz J Infect Dis.* 2010;14:19-23.
28. Mateos ML, Chacon de Antonio J, Rodriguez-Dominguez M, Sanz I, Rubio MD. Evaluation of a prototype real-time PCR assay for the separate detection of human papilloma virus genotypes 16 and 18 and other high risk human papillomavirus in cervical cancer screening. *Enferm Infecc Microbiol Clin.* 2011;29:411-4.
29. Heideman DA, Hesselink AT, Berkhof J, et al. Clinical validation of the cobas 4800 HPV test for cervical screening purposes. *J Clin Microbiol.* 2011;49:3983-5.
30. Martin IW, Steinmetz HB, Lefferts CL, Dumont LJ, Tafe LJ, Tsongalis GJ. Evaluation of the cobas 4800 HPV test for detecting high-risk human papilloma-virus in cervical cytology specimens. *Pathogens.* 2012;1:30-6.
31. Szarewski A, Mesher D, Cadman L, et al. Comparison of seven tests for high-grade cervical intraepithelial neoplasia in women with abnormal smears: the predictors 2 study. *J Clin Microbiol.* 2012;50:1867-73.
32. Meijer CJ, Berkhof J, Castle PE, et al. Guidelines for human papillomavirus DNA test requirements for primary cervical cancer screening in women 30 years and older. *Int J Cancer.* 2009;124:516-20.